

International Interdisciplinary Virtual Conference on 'Recent Advancements in Computer Science, Management and Information Technology' International Journal of Scientific Research in Computer Science, Engineering and Information Technology| ISSN : 2456-3307 (www.ijsrcseit.com)

IR, NMR Spectral Analysis of Newly Synthesized Lactose Containing Heterocyclic Compounds

Kavita M. Heda

Department of Chemistry, Shri R. L. T. College of Science, Akola 444001, Maharashtra, India

ABSTRACT

The IR spectroscopy theory utilizes the concept that molecules tend to absorb specific frequencies of light that are characteristic of the corresponding structure of the molecules. It is an essential tool for the structural analysis of newly synthesized compound. Infrared spectroscopy is the measurement of the interaction of infrared radiation with matter by absorption, emission, or reflection. It is used to study and identify chemical substances or functional groups. It is the spectroscopy that deals with the infrared region of the electromagnetic spectrum that is light with a longer wavelength and lower frequency than visible light. It covers a range of techniques, mostly based on absorption spectroscopy. Many nuclei have spin, and all nuclei are electrically charged, according to the NMR principle. An energy transfer from the base energy to a higher energy level is achievable when an external magnetic field is supplied. Nuclear Magnetic Resonance (NMR) spectroscopy is an analytical chemistry technique used in quality control and research for determining the content and purity of a sample as well as its molecular structure.

Novel 4-aryl-5-p-tolylimino-3-hepta-O-benzoyl-β-D-lactosylimino-1, 2, 4-dithiazolidines (hydrochlorides) have been synthesized by the interaction of several 1-hepta-O-benzoyl-β-D-lactosyl-3-aryl thiocarbamides with N-p-tolyl-S-chloro isothiocarbamoyl chloride. The newly synthesized compounds have been characterized by analytical and IR, 1H NMR. The polarimetric studies of the title compounds have been carried out.

Keywords: - IR, 1H NMR spectroscopy, 1, 2, 4-dithiazolidines etc.

I. INTRODUCTION

Disaccharides & Polysaccharides are important bio macromolecules with numerous beneficial functions and a wide range of industrial applications. Functions and properties of Disaccharides & Polysaccharides are closely related to their structural features. Infrared (IR) spectroscopy is a well-established technique which has been widely applied in polysaccharide structural analysis. In this paper, the principle of IR and interpretation of polysaccharide IR spectrum are briefly introduced. Classical applications of IR spectroscopy in polysaccharide structural elucidation are reviewed from qualitative and quantitative aspects. Some advanced IR techniques including integrating with mass spectrometry (MS), microscopy and computational chemistry are introduced and their applications are emphasized. These emerging techniques can considerably expand application scope

Copyright: © the author(s), publisher and licensee Technoscience Academy. This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited



of IR, thus exert a more important effect on carbohydrate characterization. Overall, this review seeks to provide a comprehensive insight to applications of IR spectroscopy in polysaccharide structural analysis and highlights the importance of advanced IR-integrating techniques.

Fundamentals of near-infrared spectroscopy in the study of carbohydrates, as well as the application of partial least squares regression (PLSR) and principal component analysis (PCA), as the most useful chemo metric techniques involved in carbohydrate analysis. The theoretical aspects and practical applications starting from simple to complex carbohydrates mixtures are covered.

For example, NMR can quantitatively analyze mixtures containing known compounds. For unknown compounds, NMR can either be used to match against spectral libraries or to infer the basic structure directly. Once the basic structure is known, NMR can be used to determine molecular conformation in solution as well as studying physical properties at the molecular level such as conformational exchange, phase changes, solubility, and diffusion. In order to achieve the desired results, a variety of NMR techniques are available.

II. RESULTS AND DISCUSSION

Herein, we report the synthesis of various 4-aryl-5-p-tolylimino-3-hepta-O-benzoyl-β-D-lactosylimino-1, 2, 4dithiazolidines (hydrochlorides) by interaction of several 1-hepta-O-benzoyl-β-D-lactosyl-3-aryl thiocarbamides with N-p-tolyl-S-chloro isothiocarbamoyl chloride. All products were crystallized from ethanol before recording the physical data. The purity of compounds was checked by TLC. The spectral analyses IR, 1H NMR of the product were observed. Optical rotation of the product was also recorded.

III. EXPERIMENTAL

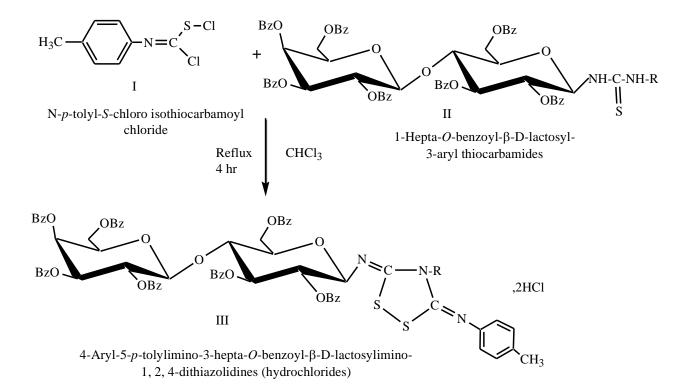
MATERIAL AND METHODS

The identities of these new N- lactocides have been established by the use of 1HNMr spectral study. ¹HNMR measurements were performed on a Bruker DRX (300MHz FT NMR) NMR spectrometer in CDCl₃ solution with TMS as internal reference 1HNMR spectra of new N-lactosides do contain some well resolved signals, including those of aromatic protons at δ 7.45-6.9ppm, lactosyl protons at δ 5.35-3.77 ppm, acetyl proton at δ 2.45-1.97 ppm, methyl protons at (Ar-CH₃) δ 1,263 ppm & other protons that are influenced by specific functionality. Similarly influence of structural features on chemical shift & coupling constant was illustrated by NMR spectra. In this way applications of NMR spectroscopy used for studies of carbohydrate are briefly considered.

Experiment No. 1: Synthesis of 4-aryl-5-p-tolylimino-3-hepta-O-benzoyl- β -D-lactosylimino-1, 2, 4-dithiazolidine (hydrochlorides) (IIIa)

To chloroform solution of *N*-*p*-tolyl *S*-chloro isothiocarbamoyl chloride (0.002M, 0.298g in 15 mL) was added chloroform solution of 1-hepta-*O*-benzoyl- β -D-lactosyl-3-aryl thiocarbamide (0.002M, 2.408g in 20mL) and the reaction mixture was refluxed over boiling water bath for 4 hr. during which evolution of HCl was noticed. The progress of reaction was monitored by TLC. After refluxing, the solvent was distilled off under reduced pressure and sticky mass obtained as residue was triturated several times with petroleum ether (60-80°C). A pale yellow solid was obtained. It was purified by chloroform-petroleum ether



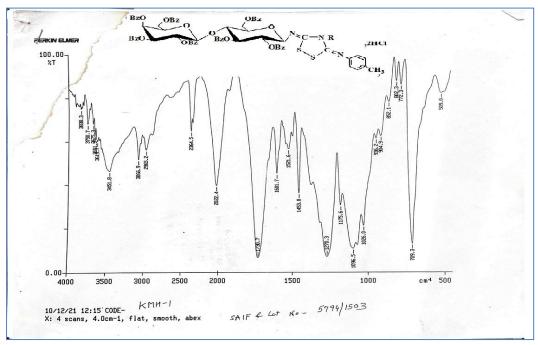


The Infrared spectral¹⁻¹⁶ analysis of the product distinctly showed bands due to Ar. C-H str., Ali. C-H str, C=O str., C=Nstr., C-N str., C-S str., S-S str., and bands due to characteristic of lactose unit.

Table 1.1: IR Spectral analysis of 4-phenyl-5- <i>p</i> -tolylimino-3-hepta- <i>O</i> -benzoyl-β-D-lactosylimino-1, 2, 4-
dithiazolidine (hydrochlorides) (IIIa)

Absorption observed (cm ⁻¹)	Assignment	Absorption expected (cm ⁻¹)
3066.9	Ar-H stretching	3080-3020 ^{9b, 10b}
2963.2	Ali. C-H stretching	2980-2920 ^{10a}
1730.7	C=O stretching	1730-1715 ^{14, 9c, 10c}
1631.7	C=N stretching	1689-147110,13-14
1270.3	C-N stretching	1400-1280 ^{10f}
1755.6	C-O stretching	1300-1041 ^{10d, 12}
1026, 936.2	characteristic of β - D- lactosyl ring	1100-1000 and 910-900 ^{12,15-16}
709.3	C-S stretching	800-680 ^{8b, 10h}

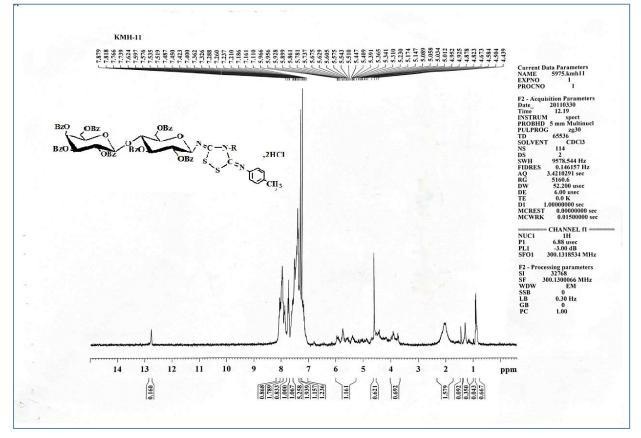




¹H NMR spectral analysis:

The ¹H NMR spectrum^{10, 14, 17-24} display the signals due to aromatic protons, lactosyl protons¹⁸⁻²⁴.

The NMR spectrum of IIIa displayed the signals due to aromatic protons at δ 8.13-7.12 ppm^{5, 14d,2-23} (44H, m, 7COC₆H₅, C₆H₅, C₆H₄) lactosyl protons at δ 6.19-3.71 ppm (14H, m)., and methyl protons (Ar-CH₃) δ 2.321ppm¹⁵⁻¹⁸ (3H, s, CH₃).



IV. CONCLUSION

In this research work, the characterizations of newly synthesized products were established on the basis of IR, ¹H NMR, spectral studies. Various lactose containing Heterocyclic Compounds were synthesized and yield of product ranged from 60-75%.

V. ACKNOWLEDGEMENT

Authors thank the Sophisticated Analytical Instrumentation Facility Panjab University, Chandigarh for providing spectral data. They are also thankful to Principal Dr. V. D. Nanoty for encouragement and necessary facilities.

VI. REFERENCES

- J. Fuentes Mota, J. M. Gracia Fernandez, C. Ortiz Mek=llet, M. A. Pradera Adrian and R. Babiano, Carbohydr. Res., 188C, 35-44 (1989)
- [2]. Mei-Sze Chua, Dong-Fang Shi, Samantha Wrigley, Tracey D. Bradshaw, Ian Hutchinson, P. Nicholas Shaw, David A. Barrett, Lesley A. Stanley, Malcolm and F. G. Steven, J. Med. Chem., 42(3), 381-392 (1999).
- [3]. Irena aleta, Marijsta Kralj, Marko Marjanovi, Barnimir Bertoa, Sanja Tomi, Gordana Pavlovi, Kreimir Paveli and Grace Karminski-Zamola, J. Med. Chem., 44(9), 1446-1455 (2009).
- [4]. Ian Hutchinson, Mei-Sez Chua, Helen L. Browne, Valentina Trapani, Tracey D. Bradshaw, Andrew D. Westwell, Malcolm and F. G. Stevens, J. Med. Chem., 44(9) 1446-1455 (2001).
- [5]. R. Babiano Caballero, J. Fuentes Mota and J. A. Galbis Perez, Carbohydr. Res., 118(1), 280-288 (1986).
- [6]. Margareta, Avram and G. H. Mateeseu, "Infrared Spectroscopy, Application in Organic Chemistry", John Wiley and Sons, INC, New York (1970) P (a) 293.
- [7]. C. T. Creswell, O. A. Runquist and M. M. Campbell, "Spectral Analysis of Identification of Organic Compounds". An introductorary programmed text, 2nd ed., Burgess Publishing company USA (1972).
- [8]. N. B. Colthup, L. H. Daly and S. E. Wibereley, "Introduction to Infrared and Raman Spectroscopy" Academic press, New York, 1964. P, (a)279,(b)344
- [9]. D. H. Williams and I. Fleming, "Spectrometric Methods of Organic Chemistry",4th ed., Tata-McGraw Hill (1991), P(a)40, (b) 41, (c) 47, (d) 53.
- [10]. R. M. Silverstein and F. X. Webster, "Spectrometric Identification of Organic Compounds", 6th ed., John Wiley and Sons, Inc. New York (2004), P(a) 83, (b) 86, (c) 97, (d) 98, (e) 102, (f) 103, (g) 104, (h) 106.
- [11]. H. Spedding, Adv. Carbohydr. Chem., 19, 23-49 (1964).
- [12]. K. Lohith et. al., Eur. J. Org. Chem., 41, 1059-1072 (2006).
- [13]. J. Fuentes, W. Moreda, C. Ortiz, I. Robina and C. Welsh, Tetrahedron L, 48(31), 6413-6424 (1992).
- [14]. J. R. Dyer, "Aplication of Absorption Spectroscopy of Organic Compounds",8th ed., Prentice Hall (1991), P (a) 36, (b) 37, (c) 84, (d) 88.
- [15]. R. Verma, S. Y. Kulkarni, C. I. Jose and V. S. Pansave, Carbohydr. Res., 113, 25-32 (1984).
- [16]. Zhiqun Dai, Fanqui Chetangi Wu and Wei Li, J. Chem. Research (S), 106-107 (2001).



- [17]. Alfonzo D. Jordan, Chi Luo and Allen B. Reitz, J. Org. Chem., 68(22), 8693-8696 (2003).
- [18]. R. U. Lemiexu and H. Driguez, J. Am. Chem. Soc., 97, 4069 (1975).
- [19]. A. Bax, W. Egan and P. Kovac, J. Carbohydr. Chem., 3, 593 (1984).
- [20]. Joaquin Isac-Gracia, Francisco G. Calvo-Flores, Fernamdo Hernandez-Mateo and Franscisco Santoyo-Gonzalez, Eur. J. Org. Chem., 383-390 (2001).
- [21]. Jose, L. B. Balnco, C. S. Barria, J. M. Benito, C. O. Mellet, J. Fuentes, F. S. Gonazalez and Jose M. Gracia, Synthesis, 11, 1907-1914, (1995).
- [22]. S. A. Barker, J. Homer, M. C. Keith and L. F. Thomas, J. Chem. Soc., 538 (1963).
- [23]. Norma B. D. Accorsa, and Inge M. E. Thiel, Carbohydr. Res., 124, 177-184 (1983).
- [24]. F. Sztaricskai, G. Pinter, E. Roth, P. Herczegh, S. Kardos, F. Rozgonyi and Z. Boda, J. Antibiot., 60(8), 529-533 (2007).

